

Trichoptera as bioindicators of habitat integrity in the Pindaíba river basin, Mato Grosso (Central Brazil)

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Abstract – We evaluated the influence of environmental integrity and the potential as bioindicator of larval stages of species of Trichoptera in 20 streams of first to fourth order in the Pindaíba river basin, Mato Grosso, Central Brazil. We measured stream integrity with the habitat integrity index (HII), establishing three levels of conservation: preserved, altered and impacted environments. We used (i) simple regression to assess the effect of habitat integrity on species abundance of Trichoptera and (ii) the indicator species analysis (IndVal) to assess the potential as bioindicator of each species. We found that 12 morphospecies showed relationship with HII: six species were bioindicators of preserved and two species of altered environments. Morphospecies that showed relationship in the two analyses (i and ii) were considered strong bioindicators, considering that the other species supported higher environmental variation, becoming evident that loss of physical structure reduces the abundance of organisms specialized in preserved environments. The results showed that the distribution and abundance of trichopterans can be an indicator of habitat integrity. Trichopteran species have bioindicator potential, corroborating the hypotheses of this work that abundance of organisms will be smaller in environments with low integrity, and that many species are specific to preserved environments, disappearing from impacted environments, and also characteristic species of altered environments.

Key words: Environmental conservation / macroinvertebrates / ecological integrity / Cerrado

Introduction

Habitat loss changes the biological structure of rivers, directly affecting the composition of aquatic macroinvertebrate and their food resources (Bispo *et al.*, 2006; Nessimian *et al.*, 2008; Dias-Silva *et al.*, 2010). The abundance of some species varies with changes in environmental integrity and even reaching extinction-risk levels (Angermeier and Karr, 1994). Such perturbations in water bodies can exclude specialist species, opening room to the colonization of generalist species (Bahar *et al.*, 2008).

The composition of macroinvertebrates in streams can be used as a parameter to measure environmental conditions (Lammert and Allan, 1999). Species that respond to environmental changes are known as bioindicators, and are very useful in environmental assessment

programs (Niemi and McDonald, 2004; Bonada *et al.*, 2006). Recent studies (*e.g.*, Cortezzi *et al.*, 2009; Utz *et al.*, 2009) showed that a variety of macroinvertebrates can be used as aquatic indicators in sites with agricultural or urban activities.

Some studies demonstrated the effects of anthropic disturbances on the structure and composition of aquatic insects in eastern Mato Grosso state. For example, the environmental integrity and complexity strongly affect the community of larval Odonata (Juen *et al.*, 2007) and a richness of Gerromorpha (Dias-Silva *et al.*, 2010), and Souza *et al.* (2010) observed that physical changes were the main causes of disruption of beta diversity of Baetidae. Shimano *et al.* (2010) considered that the removal of gallery forest and dams caused the species loss and decrease in abundance of Ephemeroptera.

Trichopterans along with ephemeropterans and plecopterans are the groups of aquatic insect most used in

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programs of aquatic environment assessment, mainly due to their high richness, ecological diversity and abundance in several types of aquatic habitats (Rosenberg and Resh, 1993; Flint *et al.*, 1999), especially in the lotic systems (Holzenthal *et al.*, 2007). These groups also are important in the trophic dynamics and energy flow in lakes, rivers and streams (Wiggins, 1996).

Trichopterans are very sensitive to changes in physical and chemical structure of aquatic environments (Spies *et al.*, 2006), anthropogenic alteration and variation in canopy cover. Accordingly, these are also the main variables affecting its distribution in the Cerrado (Oliveira and Froehlich, 1997). Additionally, several variables can increase their vulnerability to climatic and environmental changes and extinction risk, such as limited distribution, narrow ecological niche and low dispersal habitability (Hering *et al.*, 2009).

The goal of this study was to evaluate if the distribution and abundance of trichopterans reflect changes in the integrity of streams in eastern Mato Grosso, central Brazil, and select bioindicator species of habitat integrity.

Material and methods

Study site

The study was carried out in the Pindaíba river basin, eastern Mato Grosso, central Brazil (between 14°16'46" and 15°57'17"S; 51°26'37" and 52°37'03"W). We sampled five streams: Taquaral (TS), Papagaio (PS), Cachoeirinha (CS), da Mata (MS) and Caveira (CVS; Fig. 1). The Pindaíba river basin is a tributary of das Mortes river, with approximately 10 029 km² of area. The predominant economic activity in this region is extensive livestock.

The regional climate is classified as *Cwa* according to Köppen classification (Brasil, 1981), with two well-defined seasons: a dry period, from May to September, and a rainy period, from December to March. The mean annual rainfall varies from 1200 to 1600 mm and mean temperature varies about 20–25 °C, with September and October the warmest months.

Data sampling

The sampling was taken in first-, second-, third- and fourth-order streams (following Strahler, 1957), with a total of 20 sampling sites. Samplings were carried out in TS, PS and CS streams during 2005, and between 2007 and 2008 in MS and CVS streams. To reduce the effect that seasonal variation in the community of Trichoptera, samples were collected in three seasons: rainy season (January), dry (July–August) and beginning of the rainy season (from September and November), with a total of 60 samples. Due to the high variability of climatic changes during the year, we assumed that there was no autocorrelation of residuals due to temporal pseudo-replication (Hurlbert, 1984).

We collected trichopteran larvae with a 18-cm diameter sieve and 250 µm mesh, mounted on dip-net, based on training the net over the bottom, in 100-m linear transects, subdivided into 20 stretches of 5 m each (after Ferreira-Peruquetti and De Marco, 2002). We collected the substrate at the stream margin three times in each stretches.

We used several identification keys to species and morphospecies of Trichoptera (Angrisano, 1995; Wiggins, 1996; Holzenthal and Pes, 2004; Pes *et al.*, 2005, 2008). Specimens were preserved in 85% alcohol and are housed at the Coleção Zoobotânica James Alexander Ratter (CZNX), Universidade Estadual do Mato Grosso, Nova Xavantina, MT, Brazil. The number of morphotypes follows the records in this collection.

We estimated the environmental integrity of streams using the habitat integrity index (HII) (Nessimian *et al.*, 2008). This index is based on the measurement of several visually assessed parameters related to land use, riparian zone conservation, streambed characteristics, and characteristics of stream channel morphology related to margin and streamlet bed structure. Then, the index is directly related to the degree of environmental conservation and has been successfully used in other studies to evaluate the integrity in aquatic systems (Dias-Silva *et al.*, 2010; Shimano *et al.*, 2010; Juen and De Marco, 2011; Nogueira *et al.*, 2011; Souza *et al.*, 2011).

We initially calculated the HII in an ordinal scale and later converted it to a continuous scale from 0 to 1. Subsequently, we used the quantitative HII to establish three conservation categories: preserved, altered and impacted environments, according to field observations. Lastly, we used these categories to identify indicator organisms of environmental integrity.

Conservation categories

Impacted environments were sites with HII between 0.51 and 0.66, with a total of eight sites (CS_1, CS_4, CVS_1, CVS_4, PS_2 and TS_4). Due to heavy agricultural mechanization, the gallery forest at both margins of these streams are highly deforested, replaced by pastures or soy monoculture, cattle raising. Other processes, such as erosion and silting are also observed. Since stream margins are exposed, the river channel lacks retention mechanisms and consequently little deposited organic material. This situation is even worse in CRC_1, an intermittent stream in a flat area, with high cattle trampling, and the entire CVS stream, dammed at several places, becoming a semi-lotic environment.

Altered environments had HII between 0.69 and 0.78, and corresponded to five sites (CS_2, CS_3, PS_3, PS_4 and TS_3). Environmental changes measured in this study refer mainly to changes due to partial removal of marginal vegetation, cattle raising, or farming activities. At these points, part of the gallery forest was maintained at least in one of the margins and beyond the riverside zone. Furthermore, there are Cerrado patches among pasture areas, riparian vegetation varied between 5 and 30 m in



Fig. 1. Sampling sites in the Pindaíba river basin, Mato Grosso, Brazil. CS: Cachoeirinha stream; CVS: Caveira stream; MS: da Mata stream; PS: Papagaio stream; TS: Taquaral stream. The numbers 1, 2, 3 or 4 indicate streams orders.

width, there was retention mechanisms in the water course, such as stones and trunks, stable banks with some cutting, riverbed with silt, gravel and sand, and in some sites, leaf detritus and woody material with sediments.

Preserved environments had HII varying from 0.82 to 0.96, with a total of seven sites (MS_1–MS_4, PS_1, TS_1 and TS_2). At these points, the predominant characteristics were: more than 50 m in width of gallery forest, continuously with the adjacent forest, retention mechanisms strongly fixed, lack of banks, little or no

accumulation of sediments, riverbed, with stones grouped together, mosses and algae patches, and leaf detritus and woody material without sediment.

Data analysis

We used a canonical correspondence analysis (CCA; Ter Braak, 1986) to test for a relationship between environmental variables and morphospecies of Trichoptera.

We used the species abundance in each stream and the matrix of environmental variables with physico-chemical HII, pH, conductivity, water hardness, width and depth of the channel. We used only species with 10 or more individuals [see Appendix 1 (available in the online version) for species abundance used in CCA]. This procedure is considered appropriate in ordination techniques in general, since rarity or low density increases the volume of calculations, causing interpretation errors. The removal of rare species does not affect the results in a relevant way. After a preliminary analysis, we eliminated turbidity, dissolved oxygen, phosphorus and water temperature, because they had a low correlation with the ordination axes and the analysis was rerun. The biotic and environmental factors (except pH) were log-transformed in the case of the abundance matrix $\log(x + 1)$.

To test for the significance of the CCA axes, we used a Monte Carlo permutation test (Ter Braak, 1986). Analyses were performed in the R environment (R Development Core Team, 2010).

To evaluate the effect of environmental integrity on trichopteran, we used a simple linear regression (Zar, 1999). The independent variable was the HII and the dependent variable was species/morphospecies with 10 or more individuals. The use of species with minimal abundance of 10 individuals is necessary because of rarity to be very common in aquatic communities and, thus, it would be very difficult to meet the assumptions of statistical tests.

We analyzed the bioindicator potential of trichopteran species/morphospecies using the *Indicator Value Method* (IndVal; Dufrêne and Legendre, 1997). This index measures the degree of specificity (relationship of a species with a specific variable) and the degree of species fidelity (every time that some condition was met, the species was present) in relation to some environmental category. In this study, the variables used were integrity levels of sampling sites (preserved, altered and impacted).

Results

We collected 5383 trichopteran larvae, belonging to 10 families (Appendix 1 available in the online version). We identified 19 genera and 66 species/morphospecies. The genus *Nectopsyche* was the most abundant (3732 individuals) and also the most speciose (eight species).

Among the species/morphospecies with 10 or more individuals ($N = 30$), 12 were related to HII and eight were considered indicators of environmental integrity (Table 1).

The total variance in the trichopteran larvae community determined by CCA was 3.826. The total variance or inertia indicates the total amount of variability potentially explained. The first three correlations between the biotic and abiotic dataset were 0.946, 0.917 and 0.837, respectively. The first three CCA axes accounted for 27.6% of the variation of the trichopteran larvae community. Monte Carlo simulations demonstrated that the first three axes were significant (Table 2).

CCA demonstrated that HII and pH were negatively correlated and width and depth of channel were positively correlated with the first axis. Conductivity and water hardness were positively correlated and nitrate was negatively correlated with the second axis (Table 2).

The following patterns can be described to species/morphospecies: *Chimarra* sp. 6, *Leptonema* sp. 1, *Marilia* sp. 2, *Nectopsyche* sp. 1, *Phylloicus* sp. 2, *Polycentropus* sp. 1 and *Smicridea* sp. 5 occurred in streams with high index of habitat integrity. *Chimarra* sp. 4, *Chimarra* sp. 5, *Helicopsyche* sp. 3, *Leptonema maculatum* Mosely, 1933, *Macronema* sp. 2, *Marilia* sp. 1, *Oecetis* sp. 2, *Phylloicus* sp. 5, *Smicridea* sp. 6 and *Triplectides* sp. 1 occurred in wider and deeper streams, with low HII. *Helicopsyche* sp. 4, *Notalina* sp. 1 and *Polyplectropus* sp. 2 occurred in streams with high pH. The optima of *Cernotina* sp. 1 and *Polyplectropus* sp. 4 in the second CCA axis was in the streams with high conductivity and water hardness, whereas *Macrostemum* sp. 1 and *Triplectides* sp. 2 were more abundant in the sites with high nitrate (Fig. 2). Eleven species showed positive relationship with environmental integrity (*Chimarra* sp. 6, *Helicopsyche* sp. 4, *Leptonema* sp. 1, *Marilia* sp. 1 and sp. 2, *Nectopsyche* sp. 1, *Notalina* sp. 1, *Phylloicus* sp. 2, *Polycentropus* sp. 1, *Polyplectropus* sp. 2 and sp. 4). Only *Macronema* sp. 9 was negatively related with integrity, with higher abundance in environments with less integrity.

Helicopsyche sp. 4, *Marilia* sp. 2, *Nectopsyche* sp. 1, *Notalina* sp. 1, *Phylloicus* sp. 2 and *Polycentropus* sp. 1 can be considered indicators of preserved sites (IndVal). *Helicopsyche* sp. 3 and *Macronema* sp. 2 can be considered indicators of altered sites. No species was indicators of impacted environments.

Discussion

Establishing quick and appropriate ways to measure impacts is essential to recover and preserve natural resources. In this regard, biomonitoring can efficiently assess species loss and the synergistic effects of anthropogenic activities (Rosenberg and Resh, 1993; Buss *et al.*, 2003). We found that filter-collecting morphospecies (*Chimarra* sp. 6, *Leptonema* sp. 1, *Polyplectropus* sp. 2 and *Polyplectropus* sp. 4) were positively related with environmental integrity, but they were not considered good bioindicators (Merritt and Cummins, 1984; Cummins *et al.*, 2005). Several studies postulated that these genera would be present in riffle areas, *e.g.*, associated *Polyplectropus* (Martins-Silva *et al.*, 2008), *Chimarra* (Wiggins, 1996) and *Leptonema* (Baptista *et al.*, 1998). In our study site, these genera occurred in both preserved and altered environments, but the low abundance and narrow distribution of *Chimarra* sp. 6 and *Polyplectropus* sp. 2 do not make them good indicators. The broader distribution of *Leptonema* sp. 1 and *Polyplectropus* sp. 4 in both preserved and altered environments, and the lack of fidelity to one of the two types of conservation categories may explain why these

Table 1. HII related to morphospecies (simple linear regression) and species (IndVal) bioindicator of conservation status, Pindaíba river basin, 2005 and 2007–2008 (IV: indicator value of morphospecies; A: altered; P: preserved; –: absence of indication). Numbers in bold indicate significant results.

Species/morphospecies	Regression			IndVal				Indicator
	r^2	P	Equation	IV	Mean	SD	P	
<i>Ceratotina</i> sp. 1	0.185	0.058	$y = -0.524 + 1.153x$	39.7	34.5	9.20	0.252	–
<i>Chimarra</i> sp. 4	0.016	0.596	$y = 0.490 - 0.402x$	12.2	23.6	10.87	0.914	–
<i>Chimarra</i> sp. 5	0.033	0.441	$y = 0.443 - 0.423x$	11.3	21.1	10.47	0.855	–
<i>Chimarra</i> sp. 6	0.220	0.037	$y = -0.910 + 1.412x$	28.6	16.3	9.42	0.169	–
<i>Helicopsyche</i> sp. 3	0.008	0.710	$y = 0.057 + 0.189x$	49.6	28.4	10.42	0.050	A
<i>Helicopsyche</i> sp. 4	0.355	0.006	$y = -0.844 + 1.425x$	56.6	26.8	10.60	0.021	P
<i>Leptonema maculatum</i>	0.008	0.715	$y = 0.228 - 0.191x$	9.0	16.1	9.21	0.762	–
<i>Leptonema sparsum</i>	0.077	0.238	$y = 1.000 - 0.894x$	35.3	31.5	9.60	0.287	–
<i>Leptonema</i> sp. 1	0.377	0.004	$y = -1.211 + 2.342x$	45.2	34.2	8.82	0.114	–
<i>Macronema</i> sp. 1	0.155	0.086	$y = -0.599 + 1.359x$	36.8	32.9	9.32	0.295	–
<i>Macronema</i> sp. 2	0.009	0.699	$y = 0.471 - 0.207x$	61.5	33.8	8.51	0.005	A
<i>Macronema</i> sp. 9	-0.198	0.049	$y = 0.945 - 1.085x$	30.9	20.9	10.36	0.127	–
<i>Macrostemum</i> sp. 1	0.011	0.659	$y = -0.086 + 0.376x$	12.8	21.6	10.87	0.782	–
<i>Marilia</i> sp. 1	0.279	0.017	$y = -1.157 + 2.430x$	40.7	33.8	8.82	0.187	–
<i>Marilia</i> sp. 2	0.437	0.002	$y = -1.618 + 2.697x$	57.1	26.7	10.33	0.013	P
<i>Nectopsyche</i> sp. 1	0.317	0.010	$y = -2.534 + 4.376x$	68.0	35.1	11.20	0.015	P
<i>Notalina</i> sp. 1	0.266	0.020	$y = -0.592 + 0.919x$	42.9	19.4	10.19	0.040	P
<i>Oecetis</i> sp. 2	0.024	0.511	$y = -0.059 + 0.269x$	19.4	25.1	10.49	0.639	–
<i>Phylloicus</i> sp. 2	0.735	0.000	$y = -2.501 + 4.030x$	83.5	32.1	10.25	0.000	P
<i>Phylloicus</i> sp. 6	0.016	0.593	$y = 0.403 - 0.259x$	45.4	29.6	9.64	0.078	–
<i>Polycentropus</i> sp. 1	0.415	0.002	$y = -1.679 + 2.560x$	42.9	19.0	10.36	0.038	P
<i>Polyplectropus</i> sp. 2	0.247	0.026	$y = -0.763 + 1.279x$	35.0	25.6	10.90	0.187	–
<i>Polyplectropus</i> sp. 3	0.011	0.666	$y = 0.024 + 0.1681x$	18.5	26.3	10.19	0.733	–
<i>Polyplectropus</i> sp. 4	0.254	0.023	$y = -0.875 + 1.520x$	39.8	26.8	10.54	0.110	–
<i>Smicridea</i> (<i>R.</i>) sp. 6	0.042	0.384	$y = -0.220 + 0.635x$	18.5	25.0	10.38	0.694	–
<i>Smicridea</i> (<i>S.</i>) <i>palifera</i>	0.038	0.413	$y = -0.220 + 0.669x$	28.6	26.7	10.41	0.364	–
<i>Smicridea</i> (<i>S.</i>) sp. 5	0.089	0.200	$y = -0.6316 + 1.231x$	15.1	23.4	10.74	0.759	–
<i>Smicridea</i> (<i>S.</i>) sp. 6	0.012	0.641	$y = 0.221 - 0.208x$	8.6	17.1	8.70	1.000	–
<i>Triplectides</i> sp. 1	0.012	0.648	$y = 0.025 + 0.278x$	20.6	28.7	10.45	0.778	–
<i>Triplectides</i> sp. 2	0.147	0.096	$y = -0.448 + 0.802x$	29.2	23.1	10.56	0.198	–

Table 2. Results of CCA for trichopteran collected in streams of the Pindaíba river basin, Mato Grosso, Brazil. Correlation between sample scores for an axis derived from the taxa data and the sample scores that linear combinations of the environmental variables; and ** $P < 0.01$ (Monte Carlo, 1000 permutations).

Total variance (“inertia”) in the taxa data	3.826		
	Axis I	Axis II	Axis III
Eigenvalue	0.472**	0.340**	0.245**
Variance in taxa data			
% of variance explained	12.3	8.9	6.4
Cumulative % explained	12.3	21.2	27.6
Pearson correlations, Taxon-Env	0.946**	0.917**	0.837**
“Intra set correlations”			
HII	-0.887	-0.098	0.282
pH	-0.604	0.234	-0.381
Conductivity	-0.327	0.630	-0.368
Nitrate	0.259	-0.591	0.631
Water hardness	-0.372	0.455	0.221
Width	0.508	0.38	0.291
Depth	0.639	0.111	-0.037

species were not recovered as good indicators in the IndVal analysis. Among them, only *Chimarra* sp. 6 and *Leptonema* sp. 1 were strongly associated with HII. The other genera were related to hardness and conductivity.

The abundance of *Helicopsyche* sp. 4, *Marilia* sp. 2, *Nectopsyche* sp. 1, *Notalina* sp. 1, *Phylloicus* sp. 2 and *Polycentropus* sp. 1 increased as HII increase, which makes them bioindicators of preserved environments. Except for *Helicopsyche*, which is a periphyton scraper in small streams (Merritt and Cummins, 1984; Holzenthal *et al.*, 2007) and *Polycentropus*, which is a predator, but include algae in its diet (Reiso and Brittain, 2000), all the previous species are shredders.

Previous studies (Holzenthal *et al.*, 2007; Nessimian *et al.*, 2008) found that *Helicopsyche* was related to environments of high integrity, containing stones with periphyton, an important resource for its diet. Holzenthal and Hamilton (1988) found that *Polycentropus* had a limited distribution in forested streams, mostly of clear water, rapid flow and relatively no pollution. Our data for this genus are in accordance with their results. A previous study in the same study area (Dias-Silva *et al.*, 2010) found

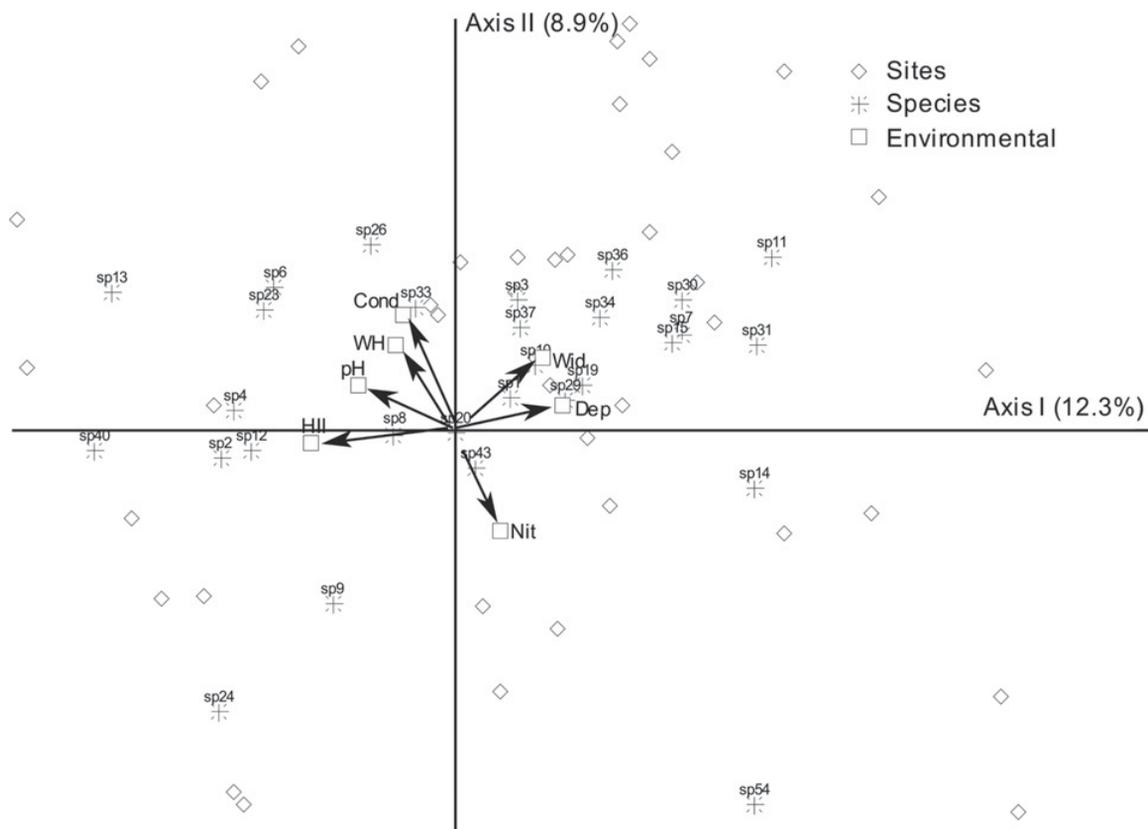


Fig. 2. Scores of CCA for trichopterans and environmental variables recorded along streams in the Pindaíba river Basin, Nova Xavantina, Mato Grosso. The abbreviations of taxa names are listed in Appendix 1 available in the online version. Habitat integrity index (HII), hydrogenionic potential (pH), conductivity (Cond), nitrate (Nit), water hardness (WH), channel width (Wid), depth of the channel (Dep).

that sites with higher integrity had higher richness of predator species of Gerromorpha.

Marilia sp. 2, a shredder species, was distributed in five preserved sites and is confirmed as potential bioindicator. Another study (Couceiro *et al.*, 2007) showed that *Marilia* was an indicator of non-impacted streams. The results also reinforce other evidence that species of *Nectopsyche* and *Phylloicus* are inhabitants of sites of high environmental heterogeneity (Crisci-Bispo *et al.*, 2007). Specifically, *Nectopsyche* species occurs mainly in riffle areas (Crisci-Bispo *et al.*, 2004) and *Phylloicus* species occurs in leaf litter in slow flowing sites, building shelter with pieces of these leaves (Pather, 2003).

Baptista *et al.* (2001) observed that *Notalina* was related to physical characteristics of upper parts of rivers, being always associated with litter deposits. In this study, *Notalina* occurred in preserved environments, but in low abundance. Conserved gallery forests are responsible for the allochthonous matter, generating detritus mainly from leaves and woody material, benefiting shredders (Silveira *et al.*, 2006).

Recovering shredders as bioindicators (HII + IndVal) indicate especially intact riparian forest with adjacent forest in first-order streams. Not surprisingly, we found the highest abundances of *Nectopsyche* sp. 1 and

Phylloicus sp. 2 in such sites. On the other hand, *Macronema* sp. 2 and *Helicopsyche* sp. 3 were little abundant in altered sites, even though they were recovered as indicator species of environments with some alteration.

Species of *Macronema* seem to be little selective, since they do not show preference for substrates (Fidelis *et al.*, 2008), but occurs commonly in roots (Flint and Bueno-Soria, 1982). This low selectivity can favor the occurrence of *Macronema* sp. 2 in altered sites. Indeed, there was an increase in abundance of *Macronema* sp. 9 as HII decreases, indicating that this is a tolerant species.

The recovering of *Helicopsyche* sp. 3 as an indicator species may be related to increase of light input as a result of reduced riparian forest, along with an increase in width and flow due to the order of altered sites, reducing shading and increasing primary production, essential for scrapers.

Our results indicate that trichopterans are highly related to habitat integrity. The six morphospecies whose abundances were influenced by HII and considered indicators of environmental conditions should be identified to the species level. The advantage of using this taxonomic level is straightforward: species of the same genus have different levels of stress tolerance, and several morphospecies of one genus may have different indicator values, distribution and abundances.

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