Three species of *Isoperla* occur in Fennoscandia. The most common is *I. grammatica*, although it has not been recorded above the subalpine vegetation belt. *I. difformis* has a similar distribution, but is less common. *I. obscura* occurs mainly at high latitudes and altitudes, although it is also abundant in large rivers.

Eggs were reared at temperatures of 4, 8, 12, 16, 20 and 24°C. *I. difformis* did not hatch at 4°C, *I. grammatica* at 4 and 8°C while *I. obscura* did not hatch at 16°C and above. Egg mortality was high in *I. difformis* at 24°C. Incubation period decreased with increase in temperature and varied between 28 and 68 days in *I. grammatica* and between 15 and 83 days in *I. difformis*. For *I. grammatica* and *I. difformis*, the relationship between the egg incubation period and the water temperature was linear on logarithmic scales. For *I. obscura*, no such relationship was found and the mean incubation period at all temperatures was 305 ± 57 (S.D.) days. Explanations are presented to explain these differences in egg incubation characteristics.

**Introduction**

Three species of *Isoperla* occur in Fennoscandia: *I. difformis*, *I. grammatica* and *I. obscura* (Brinck 1949). *I. grammatica* is the most common in Norway, occurring in all kinds of streams and rivers, but not in lakes. However, this species does not occur above the subalpine vegetation belt, situated at about 1 100 m a.s.l. in southern Norway (Lillehammer 1974). *I. difformis* has a similar distribution, but is far less common and has not been recorded above 700 m a.s.l. in northern Norway *I. obscura* is common in all kinds of freshwater environments, but in the southern parts of Norway the species is only common in high altitude localities, except for large lowland rivers. This is the only *Isoperla* species which has been recorded in the mid-alpine vegetation belt, being recorded up to about 1 500 m a.s.l. in southern Norway.

In an attempt to obtain a better understanding of the distribution and environmental strategies of Norwegian stoneflies, egg incubation and growth studies are being carried out in the laboratory on different species. Through these studies we hope to be able to categorize different types of egg development and nymphal growth patterns in stoneflies. Previous contributions to our synthesis (see Lillehammer et al., this volume) include the studies by Lillehammer 1975, 1976, Brittain 1978, 1983, Brit-l
tain et al. 1984 and Saltveit 1977, as well as these studies on Isoperla.

Temperature is a major factor influencing stonefly distribution (Lillehammer 1974). Distribution patterns of species may therefore be in part explained by their capability to adapt their life cycle strategies to different kinds of temperature regimes. Controlled laboratory studies enable us to assess development time and temperature requirement of the different life cycle stages. This paper presents data on the temperature requirements and limitations for the egg stage of I. difformis, I. grammatica and I. obscura.

1. — Methods

Adult Isoperla, along with sufficient water for egg incubation studies, were collected from the field sites, Saeterbekken (110 m a.s.l.) and Sørkedalselva (125 m a.s.l.) in the Oslo region (60° 00' N, 10° 30' E) and Valdresflya (1 400 m a.s.l.) in the Jotunheimen mountains (61° 25' N, 8° 52' E), during the emergence period. The adults were transported to the laboratory where they were kept in small plastic boxes placed in constant temperature cabinets at 10°C. On most days the boxes were transferred to 20°C for a few hours to increase the chance of mating and oviposition. Each box contained a small Petri dish with water from the field site, together with small twigs and leaves, which provided shelter. The boxes were inspected daily and any egg batches removed and counted, before being placed at constant temperatures at 4°C intervals between 4 and 24°C. All egg batches were incubated in total darkness and were inspected daily for hatching. The duration of egg incubation at the various temperatures was taken as the period in days from oviposition until 50% of the eggs that eventually hatched in any one batch had hatched.

2. — Results

In I. difformis no hatching occurred at 4°C (Table I). At 8°C the egg incubation period was about 80 days with a hatching success of 90%. The mean incubation period decreased with increasing temperature, being 15-20 days at 20-24°C. The low hatching success in some of the egg batches was due to fungal infections.

The eggs of I. grammatica did not hatch at 4 and 8°C, while the mean incubation period at 12°C was 54 days and decreased with increasing temperature (Table II). However, the differences were small between 16 and 24°C. The hatching success was high at all temperatures where hatching occurred (90-100%).

Eggs of I. obscura hatched at 4, 8 and 12°C, but not at 16 and 20°C (Table III). However, the hat-
Table III. Egg incubation data for individual egg batches from *Isoperla obscura* from Valdresflya, showing incubation temperature (T° C), number of eggs (n), days for first egg to hatch, mean (50 %) incubation period in days, duration of hatching in days and percent hatching success.

<table>
<thead>
<tr>
<th>T° C</th>
<th>n</th>
<th>First egg to hatch (days)</th>
<th>Incubation period (days)</th>
<th>Duration of hatching (days)</th>
<th>% Hatching</th>
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<tr>
<td>4</td>
<td>140</td>
<td>321</td>
<td>337</td>
<td>86</td>
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<tr>
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<td>84</td>
<td>320</td>
<td>342</td>
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<td>52.4</td>
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<tr>
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<td>63</td>
<td>310</td>
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<td>87</td>
<td>92.1</td>
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<td>146</td>
<td>321</td>
<td>357</td>
<td>75</td>
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<td>83</td>
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<td>43</td>
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<td>-</td>
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</table>

Hatching success at 12° C was very low. At 4 and 8° C a very long and similar incubation period was found, varying between 332 and 357 days at both temperatures. The highest hatching success was at 4° C.

For both *I. difformis* and *I. grammatica* the relationship between the egg incubation period (Y days) and temperature (T° C) was highly significant, respectively P < 0.001 and P < 0.01. The relationship was linear on logarithmic scales and well expressed by the equation:

\[
\log Y = \log a - b \log T
\]

or

\[
Y = a T^{-b}
\]

Values of the constants a and b are given in Fig. 1.

For *I. obscura* no significant relationship was found between the egg incubation period (Y-days) and temperature (T° C). Thus egg development seems independent of temperature in the range tested.

![Fig. 1. The relationship between egg incubation period (Y days) and water temperature (T° C) for *Isoperla difformis* and *Isoperla grammatica*. The equations for the regression lines are given for both species.](image)
3. — Discussion

Egg development in both *I. difformis* and *I. grammatica* was highly temperature dependent, with a short incubation period. A similar basic relationship between temperature and incubation period has been found for other stonefly populations, such as *Taeniopteryx nebulosa* (Brittain 1977), *Nemurella pictetii* (Brittain 1978) and *Capnia atra* (Brittain et al. 1984). The intercepts are higher in both *I. difformis* and *I. grammatica*, meaning that they require more degree days for development than the other mentioned species. These *Isoperla* species seem therefore restricted to areas which are able to provide the required number of degree days for egg development in a relatively short period.

In *I. obscura*, however, development seems independent of temperature in the range tested. The development period in *I. obscura* is at least 10-11 months, but some eggs have a longer development time. The presence of small nymphs in late August and September almost at the same time as the adult emerge (Lillehammer, unpubl. data), support the laboratory results. We are uncertain about any diapause. However, in some preliminary studies eggs taken from 12 and 16° C in January and transferred to lower temperatures did have a higher hatching success, although the incubation period did not differ from that found at 4 and 8° C. This is similar to the effect found when cooling down eggs of *Amphinemura standfussi* that did not hatch at 16 and 20° C (Saltveit 1977). However, a different effect was found in *Diura nanseni* where freezing broke egg diapause (Lillehammer 1976).

In Norway seven other species have a similar altitudinal distribution to *I. obscura* (Lillehammer 1974, 1984). Five of these species have a long egg development time during the winter (winter eggs) (Lillehammer 1976, Saltveit 1977, unpubl. data), and this includes all the four predator species (Systellognatha) occurring in the area. These species are north to north-eastern immigrants to Fennoscandia, but show differences in morphology in far northern areas. Studies of the egg development in widely separated populations is necessary in order to provide answers to these questions concerning inter- and intraspecific variation in life history patterns. It would also be interesting to compare immigration history and egg development. Another interesting comparison would be one of the egg development types with phylogenetic location of the species and genera.

Acknowledgement

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Literature cited


S.J. Saltveit & A. Lillehammer